RNA Secondary Structure Prediction

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Outline

- RNA folding
- Dynamic programming for RNA secondary structure prediction
- Covariance model for RNA structure prediction

RNA Basics

- RNA bases A,C,G,U
- Canonical Base Pairs
 - A-U
 - G-C
 - G-U

"wobble" pairing

Bases can only pair with one other base.





RNA Basics

- transfer RNA (tRNA)
- messenger RNA (mRNA)
- ribosomal RNA (rRNA)
- small interfering RNA (siRNA)
- micro RNA (miRNA)
- small nucleolar RNA (snoRNA)



http://www.genetics.wustl.edu/eddy/tRNAscan-SE/

RNA Secondary Structure



Pseudoknots

 Pseudoknots: a nucleic acid secondary structure containing at least two stem-loop structures which half of one stem is intercalated between the two halves of another stem.



Sequence Alignment as a method to determine structure

- Bases pair in order to form backbones and determine the secondary structure
- Aligning bases based on their ability to pair with each other gives an algorithmic approach to determining the optimal structure



Simple Example: Maximizing Base Pairing $S(i,j) = \max \begin{cases} S(i+1,j-1) + 1 & [\text{if } i,j \text{ base pair}] \\ S(i+1,j) \\ S(i,j-1) \\ \max_{i < k < j} S(i,k) + S(k+1,j) \end{cases}$

S(i,j) is the folding of the subsequence of the RNA strand from index i to index j which results in the highest number of base pairs

Simple Example: Maximizing Base Pairing

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 $i+1 \bullet - \bullet j - 1$

S(i+1,j-1)

Base pair at i and j

Images – Sean Eddy

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Unmatched at i



Images – Sean Eddy

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Umatched at j

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Bifurcation

Images – Sean Eddy

- Alignment Method
 - Align RNA strand to itself
 - Score increases for feasible base pairs
- Each score independent of overall structure
- Bifurcation adds extra dimension



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Initialize first two diagonal arrays to 0



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Fill in squares sweeping diagonally



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Bases cannot pair, similar to unmatched alignment



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Dynamic Programming – possible paths



Images – Sean Eddy

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Bifurcation – add values for all k



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Base Pair Maximization - Drawbacks

- Base pair maximization will not necessarily lead to the most stable structure
 - May create structure with many interior loops or hairpins which are energetically unfavorable
- Comparable to aligning sequences with scattered matches not biologically reasonable

Energy Minimization

- Thermodynamic Stability
 - Estimated using experimental techniques
 - Theory : Most Stable is the Most likely
- No Pseudknots due to algorithm limitations¹
- Uses Dynamic Programming alignment technique
- Attempts to maximize the score taking into account thermodynamics
- MFOLD and ViennaRNA



Energy Minimization Results



- Linear RNA strand folded back on itself to create secondary structure
- Circularized representation uses this requirement
 - Arcs represent base pairing

Energy Minimization Results



 All loops must have at least 3 bases in them Equivalent to having 3 base pairs between all arcs

Exception: Location where the beginning and end of RNA come together in circularized representation

Trouble with Pseudoknots



- Pseudoknots cause a breakdown in the Dynamic Programming Algorithm.
- In order to form a pseudoknot, checks must be made to ensure base is not already paired – this breaks down the recurrence relations

Energy Minimization Drawbacks

- Compute only one optimal structure
- Usual drawbacks of purely mathematical approaches
 - Similar difficulties in other algorithms
 - Protein structure
 - Exon finding

- Incorporates Similarity-based method
 - Evolution maintains sequences that are important
 - Change in sequence coincides to maintain structure through base pairs (Covariance)
 - Cross-species structure conservation example tRNA
- Manual and automated approaches have been used to identify covarying base pairs
- Models for structure based on results
 - Ordered Tree Model
 - Stochastic Context Free Grammar



Expect areas of base pairing in tRNA to be covarying between various species



Base pairing creates same stable tRNA structure in organisms



Mutation in one base yields pairing impossible and breaks down structure



Covariation ensures ability to base pair is maintained and RNA structure is conserved

Binary Tree Representation of RNA Secondary Structure

- Representation of RNA structure using Binary tree
- Nodes represent
 - Base pair if two bases are shown
 - Loop if base and "gap" (dash) are shown
- Traverse root to leaves, from left to right
- Pseudoknots still not represented
- Tree does not permit varying sequences
 - Mismatches
 - Insertions & Deletions



Covariance Model

- HMM which permits flexible alignment to an RNA structure
 - emission and transition probabilities
- Model trees based on finite number of states
 - Match states sequence conforms to the model:
 - MATP State in which bases are paired in the model and sequence
 - MATL & MATR State in which either right or left bulges in the sequence and the model
 - Deletion State in which there is deletion in the sequence when compared to the model
 - Insertion State in which there is an insertion relative to model
- Transitions have probabilities
 - Varying probability Enter insertion, remain in current state, etc
 - Bifurcation no probability, describes path



- Calculate the probability score of aligning RNA to CM
- Three dimensional matrix O(n³)
 - Align sequence to given subtrees in CM
 - For each subsequence calculate all possible states
- Subtrees evolve from Bifurcations
 - For simplicity Left singlet is default



- •For each calculation take into account the
 - •Transition (T) to next state
 - •Emission probability (P) in the state as determined by training data

 $S_{i,j,y}(y = MATP) = \max_{y_{next}} [S_{i+1,j-1,y_{next}} + \log \mathcal{T}(y_{next} \mid y) + \log \mathcal{P}(x_i, x_j \mid y)]$



- •For each calculation take into account the
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$$S_{i,j,y}(y = MATR, INSR) = \max_{y_{next}} [S_{i,j-1,y_{next}} + \log \mathcal{T}(y_{next} \mid y) + \log \mathcal{P}(x_j \mid y)]$$



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Images – Eddy et al.

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 - •Emission probability (P) in the state as determined by training data

Deletion – does not have an emission probability (P) associated with it

$$S_{i,j,y}(y = DEL) = \max_{y_{next}} [S_{i,j,y_{next}} + \log \mathcal{T}(y_{next} \mid y)]$$



Images – Eddy et al.

- •For each calculation take into account the
 - •Transition (T) to next state
 - •Emission probability (P) in the state as determined by training data

Bifurcation – does not have a probability associated with the state

$$S_{i,j,y}(y = BIFURC) = \max_{i-1 < =mid < =j} [S_{i,mid,y_{ieft}} + S_{mid+1,j,y_{right}}]$$

Model Training



Covariance Model (CM) Training Algorithm

• S(i,j) = Score at indices i and j in RNA when aligned to the Covariance Model

$$S(i,j) = \max \begin{cases} S(i+1,j-1) + M(i,j) \\ S(i+1,j) \\ S(i,j-1) \\ \max_{i < k < j} S(i,k) + S(k+1,j) \end{cases}$$

$$M_{i,j} = \sum_{x_i, x_j} f_{x_i x_j} \log_2 \frac{f_{x_i x_j}}{\int_{x_i} f_{x_j}} \qquad \text{Independent frequency of seeing the symbols (A, C, G, T) in locations i or depending on symbol.}$$

• Frequencies obtained by aligning model to "training model to "traing to the training model to "training model to "training model to "training model to "training model to "traing to the training model to "training model to "training to

- data" consists of sample sequences
- Reflect values which optimize alignment of sequences to model

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Mutual information for RNA Secondary Structure Prediction



GGGCUUGUAGCUCAGCU-GGU--AGAGCGCCGCCUU RECEAGEC ICC'AL GCGGUUGUG GCCAA CC A GCCCCCAUCGUCUA GGACAC GGCCU GGU -GGUU-AUGGCAUC UUCGUGGUC UAG CACCAGO UG GC

Covariance Model Drawbacks

- Needs to be well trained
- Not suitable for searches of large RNA
 - Structural complexity of large RNA cannot be modeled
 - Runtime
 - Memory requirements

References

• <u>How Do RNA Folding Algorithms Work?</u>. S.R. Eddy. <u>*Nature Biotechnology*</u>, 22:1457-1458, 2004.